Introduction to Stochastic Simulation with the Gillespie Method

David Karig April 18, 2005

Stochastic Systems



- Many systems driven by random, discrete interactions
- Traditional deterministic models may not accurately describe such systems

Example: The Lambda Switch

Virus Decision Dictated by Noise



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Outline

- Deterministic rate reaction model
- Gillespie method
- Examples
 - Lambda phage
 - Epidemiology
- Optimizations

Deterministic Model



 Given initial conditions, integrate the coupled equations for some period of time

Problem Statement



If we start with N species which can interact through one of M reactions at a given time, what will be the population levels of species after a given period of time?

Deterministic Solution



- Continuous
- Average kinetic rates represent reaction probabilities

Validity of Deterministic Solution

 $\mathsf{A} + \mathsf{B} \xrightarrow{} \mathsf{C}$







Validity of Deterministic Solution





Reaction Probabilities

- c_μ dt = average probability that a *particular* combination of reactants will react according to R_μ in the next time interval dt
- h_{μ} = number of reactant combinations
- $h_{\mu}c_{\mu} dt = a_{\mu} dt = average probability that an R_{\mu}$ reaction will occur somewhere inside V in the next time interval dt

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R₁: A + B \rightarrow C X molecules of A Y molecules of B $h_1 = XY$ reactant combinations

 $XYc_1 dt =$ probability that an R_1 reaction will occur somewhere inside V in the next time interval dt

Exact Stochastic Simulation

- Avoid averaging assumptions
- Probabilistic formulation
 - When does next reaction occur?
 - Which reaction occurs next?
- Reaction probability density function:

 $P(\tau,\mu)d\tau = probability at time t that the next reaction$ $is R_u and occurs in interval (t+<math>\tau$,t+ τ +d τ)



System State:

		t	t+τ
	А	1	2
	В	1	2
	С	8	7

Deriving Reaction PDF



 $P(\tau,\mu)d\tau = P_0(\tau) \underset{\text{probability that reaction } \mu \text{ occurs during } (t+\tau, t+\tau+d\tau)$

Deriving $P_0(\tau)$ for Reaction PDF



 Probability that none of the reactions occur in any of the K subintervals:

$$P_0(\tau) = \left[1 - \sum_{i=1}^M h_i c_i \varepsilon\right]^K = \lim_{K \to \infty} \left[1 - \frac{\sum_{i=1}^M h_i c_i \tau}{K}\right]^K = e^{-\sum_{i=1}^M h_i c_i \tau}$$

Direct Method for Generating $P(\tau,\mu)$

$$P(\tau,\mu) = h_{\mu}c_{\mu}P_{0}(\tau) = h_{\mu}c_{\mu}e^{-\sum_{i=1}^{M}h_{i}c_{i}\tau}$$
$$= a_{\mu}e^{-a_{0}\tau}$$
$$= \left(a_{0}e^{-a_{0}\tau}\right)\left(\frac{a_{\mu}}{a_{0}}\right) = P(\tau) \cdot P(\mu|\tau)$$

whenwhichnextreactionreactionoccursoccurs



 $\mathbf{a}_0 = \sum_{\mu=1}^M a_\mu$

Direct Method for Generating $P(\tau,\mu)$



 $\tau = (1/a_0) \ln(1/rand_1)$

 μ is the integer for which $\sum_{i=1}^{\mu-1} a_i < rand_2 a_o < \sum_{i=1}^{\mu} a_i$

Stochastic Simulation Algorithm

- 1. Initialization
 - Set values of c_{μ} for the M reactions.
 - Set initial population sizes
- 2. Calculate the M values a_{μ} and $a_0 = \Sigma a_{\mu}$.
- 3. Generate (τ, μ) based on $P(\tau, \mu)$
- 4. Adjust population levels according to the reaction R_{μ} , and increase t by τ
- 5. Return to Step 2

Lambda Phage Developmental Pathway



- Regulatory circuit exploits stochastic noise to produce different outcomes
- Stochastic model can predict statistics of regulatory outcomes

Lambda Phage: Regulatory Circuit



- CI and Cro competitively bind O_{R1} , O_{R2} , O_{R3}
- Cro represses P_R and P_{RM}
- CI represses P_R, can activate P_{RM}

Lambda Phage: Regulatory Circuit



- CI and Cro competitively bind O_{R1}, O_{R2}, O_{R3}
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Time Evolution of Two Runs



Epidemiology Example



Evaluation of Stochastic Simulation

• Advantages

- continuous time, discrete population changes
- captures effects of noise
- simple implementation
- small memory requirements
- Disadvantages
 - CPU intensive
 - typically must simulate many runs
 - must use good random number generator
 - periodicity affects size of simulation
 - resolution limits range of probabilities

Computational Requirements

- Memory (N + 2M + 1)
 - N species populations
 - -c and *a* values for each of M reactions; a_0
- Total time scales with number of reactions that occur
- Operations per reaction:



Optimized Direct Method (ODM)

Y. Cao, H. Li, L. Petzold. 2004. J. Chem. Phys. 121:4059-4067

- Reduce cost of searching for index μ
- Observation: Reactions are typically multiscale in a large system. Subset will frequently occur.
- Sort index of reactions based on how often they occur

$$\sum\nolimits_{i=1}^{\mu-1} a_i < rand_2 a_o < \sum\nolimits_{i=1}^{\mu} a_i$$



Optimized Direct Method (ODM)

- Reduce cost of calculating all a_μ
- Dependency graph
- Reduce cost of summing all a_{μ} to calculate a_0
- Modify a₀ by subtracting old values, adding new



Gibson and Bruck, 2000

Conclusion

- Provides means of studying role of noise in complex systems
- Can predict statistics (Lambda phage)
- Can depict behavior that deterministic simulations do not capture (epidemiology example)
- Enhancing performance an active area of investigation

Useful References / Further Study

• Implementations

- STOCKS: http://www.sysbio.pl/stocks/stocks1.html
- BioNetS: https://users.biospice.org/toolsumm.php?id=2
- http://www.staff.ncl.ac.uk/d.j.wilkinson/software/

Algorithms/optimizations

- D.T. Gillespie. A General Method for Numerically Simulating the Stochastic Time Evolution of Coupled Chemical Reactions. 1976. *J Comput Phys* 22:403-434.
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- M.A. Gibson and J. Bruck. 2000. Efficient Exact Stochastic Simulation of Chemical Systems with Many Species and Many Channels. J Phys Chem 104:1876-1889
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- A. Chatterjee and D.G. Vlachos. Binomial distribution based τ -leap accelerated stochastic simulation. 2005. *J Chem Phys* 122:024112

• Example Uses

- A. Arkin, J. Ross, H. McAdams. Stochastic Kinetic Analysis of Developmental Pathway Bifurcation in λ Phage-Infected *Escherichia coli* Cells. 1998. *Genetics* 149:1633-1648
- J. Dushoff, J.B. Plotkin, S.A. Levin, D.J.D. Earn. Dynamical resonance can account for seasonality of influenza epidemics. 2004 *PNAS*.
- S. Hooshangi, S. Thiberge, R. Weiss. Ultrasensitivity and noise propagation in a synthetic transcriptional cascade. 2005. PNAS 102:3581-3586

Acknowledgements

- Ron Weiss and the Weiss group
- PICASso
 - Steven Kleinstein
 - J.P. Singh



Comparing to Experimental Results

- Compare predicted percent lysogenization to experimental results at different infection levels
- Average phage input (API) = ratio of phage particles to cells at time of infection
- Multiplicity of infection (MOI) = phage particles per cell
- Poisson probability that a given cell will be infected with MOI=M when API=A

$$P(M,A) = \frac{A^M}{A!}e^{-A}$$

• Expected fraction of lysogens:

$$F_{lysogens}(A) = \sum_{M} P(M, A) \cdot F(M)$$

Comparing to Experimental Results



Example (deterministic assumptions)



Average rate of R_1 in dt per unit volume:

$$\frac{\left\langle X_1 X_2 c_1 \right\rangle}{V} = \frac{\left\langle X_1 X_2 \right\rangle c_1}{V}$$

Using concentrations $x_i = X_i/V$ and dividing by density of reactants:

$$k_{1} = \frac{\langle x_{1} x_{2} \rangle c_{1} V}{\langle x_{1} \rangle \langle x_{2} \rangle}$$

Deterministic assumption:

$$k_1 = c_1 V$$

Master equation

$$\frac{\partial}{\partial t} P(X_1, \dots, X_N; t) = \sum_{\mu=1}^M \left[B_\mu - a_\mu P(X_1, \dots, X_N; t) \right]$$

- B_μ dt = probability that single R reaction brings us to state X₁,...,X_N
- $a_{\mu} dt = h_{\mu}c_{\mu} dt$, where h_{μ} is the number of reactant combinations
- Often difficult to solve analytically and even numerically
- →Instead simulate individual trajectories using Monte Carlo algorithm